**SPIROPA Cruise Planning 2019**

Draft 12/3/18

May cruise – R/V *Ronald H. Brown* (30 berths)

Load: 06 May 2019

Dep: 08 May

Arr: 21 May

Unld: 22 May

July cruise – R/V *Thomas G. Thompson* (36 berths)

Load: 03 Jul 2019

Dep: 05 Jul

Arr: 18 Jul

Unld: 19 Jul

**1. Cruise Overview**

The continental shelfbreak of the Middle Atlantic Bight supports a productive and diverse ecosystem. Current paradigms suggest that this productivity is driven by several upwelling mechanisms at the shelfbreak front. This upwelling supplies nutrients that stimulate primary production by phytoplankton, which in turn leads to enhanced production at higher trophic levels. Although local enhancement of phytoplankton biomass has been observed in some synoptic measurements, such a feature is curiously absent from time-averaged measurements, both remotely sensed and *in situ*. Why would there not be a mean enhancement in phytoplankton biomass as a result of the upwelling? One hypothesis is that grazing by zooplankton prevents accumulation of biomass on seasonal and longer time scales, transferring the excess production to higher trophic levels and thereby contributing to the overall productivity of the ecosystem. However, another possibility is that the net impact of these highly intermittent processes is not adequately represented in long-term means of the observations, because of the relatively low resolution of the *in situ* data and the fact that the frontal enhancement can take place below the depth observable by satellite.

A unique opportunity to test these hypotheses has arisen with deployment of the Ocean Observatories Initiative (OOI) Pioneer Array south of New England. The combination of moored instrumentation and mobile assets (gliders, AUVs) will facilitate observations of the frontal system with unprecedented spatial and temporal resolution (Fig. 1). This will provide an ideal four-dimensional (space-time) context in which to conduct a detailed study of frontal dynamics and plankton communities.

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|  | Fig. 1. Tracks of Pioneer Array gliders (grey, magenta lines), 17 Apr – 30 Jun 2014. Red line is a cross-shelf transect on 25-26 Apr; the black triangle, diamond, and circle indicate the positions of the foot, jet and surface expression of the front, respectively. Mooring locations are shown as stars, with the central offshore mooring filled in black. Shipboard transects indicated with blue circles. The solid black boundary depicts our model domain. |

With support from NSF’s Physical, Biological, and Chemical Oceanography programs, we will undertake a set of three cruises to obtain cross-shelf sections of physical, chemical, and biological properties within the Pioneer Array. Nutrient distributions will be assayed together with hydrography to detect the signature of frontal upwelling and associated nutrient supply. We expect that enhanced nutrient supply will lead to changes in the phytoplankton assemblage, which will be quantified with conventional flow cytometry, imaging flow cytometry (Imaging FlowCytobot, IFCB), *in situ* optical imaging (Video Plankton Recorder, VPR), traditional microscopic methods, and HPLC pigments. Zooplankton will be measured in size classes ranging from micro- to mesozooplankton with the IFCB and VPR, respectively, and also with microscopic analysis. Biological responses to upwelling will be assessed by measuring rates of primary productivity, zooplankton grazing, and net community production. These observations will be synthesized in the context of a coupled physical-biological model to test the two hypotheses that can potentially explain prior observations: (1) grazer-mediated control and (2) undersampling. Hindcast simulations will also be used to diagnose the relative importance of the various mechanisms of upwelling.

Our observational plan consists of cross-frontal transects and rate measurements, conducted in a daily cycle of activity (Fig. 2). Each day will begin with determining the precise location of the front from a combination of data from the Pioneer Array, cruise observations, and remote sensing images. Rate measurements (14C and grazing incubations) will be strategically located in one of the three key regimes: inshore, offshore, and at the front. Twelve repetitions of the observational cycle (see below) will permit four replicates in each of the three regimes, facilitating estimates of the mean and variance for each. Each of the 12 cross-frontal transects will consists of a 12-station subset of the range of possible station locations shown in Fig. 2. Each specific 12-station subset will be centered on the front, essentially shifting northward or southward as movement of the front dictates. Station spacing is 7km.

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| Fig. 2. Daily sampling cycle (left) and summary of measurements (right). |

**Sampling**

CTD/VPR Transects

Underway ADCP, TSG, benchtop IFCB, benchtop FCM (Attune NxT), EIMS

Smith 3 14C incubations per day: dawn, noon, pre-sunset

Turner incubations coupled once per day; MOCNESS net tows at that station

VPR II transects

Philosophy: while each group has their own specific measurements for which they will be responsible, what makes this study so special is the synergy among the diverse set of observations. We have had to make compromises in staffing to fit our science party into the maximum of 22 berths, and as such not every group was able to bring along all the people they would have liked to have had participate. As such, we need to help each other out—there is no one on the cruise who is solely “doing their own thing.” Please keep this in mind as we try to balance the workload to accommodate all of the various sampling needs.

**Water budget (mL)**

\*TOI, O2/Ar 750 Stanley 300 all surface, limited profiles at 25% of stations

DIC 350 Smith

14C 350 Smith 3 per day

POC 2000 Smith

IFCB 100 Sosik surface, Chl max, plus 3 6-depth profiles (shelf,

front, slope) every other transect

FCM 50 Sosik same as IFCB

Chl 700 Sosik every other transect; deep samples with low F omitted

HPLC 1300 Sosik surface, Chl max, plus 3 6-depth profiles (shelf,

front, slope) every other transect

Nutrients 150 McG all depths, every other transect

Total 5760

\*Gases always sampled first

Water sample inventory: daily transects (12) \* 12 stations \* 15 depths = 2160 discrete samples

Nutrients – 1080

TOI, O2/Ar – 300

Suggested sample vial labeling: C for cast number, N for Niskin number, and include cruise. Example: Cast 54, Niskin 11, Cruise AR-29; label would be “C54 N11 AR-29”.

**Sampling depths – 15 total**

Fixed: Sfc, 10, 20, 30, 40, 60, 80, 100, 120, 150, 200, 250, 300

Floating: chl max, 10mab

**Shifts – AR29**

Noon to Midnight Midnight to Noon

CTD Operators; help with CTD prep and water sampling

Charlie Gordon

IFCB, FCM, Chl, HPLC, nutrients

Heidi (watch chief) Taylor\* (watch chief)

Kevin (nutrients) Bethany

Jackie Meredith

**Addt’l hand below** Emily (nutrients)

Gas Samplers

Zoe\*(recovery only) Zuchuan

Additional hands

1200-1600 Phil\* (also help with sampling)

1600-2000 Josh\*, Melissa C.

2000-2200 Beth\* (also help with water sampling)

\*CTD Launch and recovery

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14C – daylight only – Walker

Zooplankton – day shifts

Jeff, Chrissy, Beth, Melissa C.

VPR, MOCNESS – day shifts

Phil, Josh, Melissa P.

Media – 24/7

Dan, Saskia

**Personnel from AR29**

1. Dennis McGillicuddy
2. Phil Alatalo
3. Josh Eaton
4. Melissa Patrician
5. Zuchuan Li
6. Heidi Sosik
7. Taylor Crockford
8. Bethany Fowler
9. Kevin Archibald
10. Gordon Zhang
11. Zhen Cheng (Charlie)
12. Walker Smith
13. Jackie Friedman (WOS Lab)
14. Meredith Meyer (WOS Lab)
15. Jeff Turner
16. Chrissy Petitpas
17. Elizabeth (Beth) Larson (JT Lab)
18. Melissa Campbell (JT Lab)
19. Zoe Sandwith (Stanley lab)
20. Dan Brinkhaus – Science Media
21. Saskia Madlener – Science Media
22. Emily Shimada – MATE (Marine Advanced Technology Education) intern

**Personnel –** R/V *Ron Brown* (30 berths)

1. Dennis McGillicuddy
2. Phil Alatalo
3. Andrew Hirzel
4. TBA – McG
5. TBA – McG
6. Gordon Zhang
7. Hilde Oliver
8. Walker Smith
9. Meredith Meyer
10. Mar Arroyo
11. TBA – Smith
12. Jeff Turner
13. Chrissy Petitpas
14. TBA – Turner
15. TBA – Turner
16. TBA - Turner

**Personnel –** R/V *Thomas G. Thompson*(36 berths)

1. Dennis McGillicuddy
2. Phil Alatalo
3. Andrew Hirzel
4. TBA – McG
5. TBA – McG
6. Gordon Zhang
7. Hilde Oliver
8. Walker Smith
9. Meredith Meyer
10. Mar Arroyo
11. TBA – Smith
12. Jeff Turner
13. Chrissy Petitpas
14. TBA – Turner
15. TBA – Turner
16. TBA - Turner
17. Dan Brinkhaus

**Zhang group tasks**

1. CTD deployment, real-time CTD data processing

2. EK80 system monitoring, adjusting and data interpretation

3. Shipboard ADCP system monitoring and data interpretation

4. Real-time satellite SST data access and processing

5. Real-time Pioneer Array data access and processing

6. Real-time Wilkin model data access and processing

7. Real-time data integration and identification of the shelf break front

8. Inform decisions on the locations of CTD and biological sampling stations

**Event logger**

<http://elog-dev.whoi.edu/default/eloghelp_R2R_EventLogger_User_Guide.html>

**Retrospective data analysis**

AR16 Science Verification Cruise

AR22 Joint Program Cruise

Pioneer turnarounds

Fishing effort

**Synergy with NES LTER** <https://nes-lter.whoi.edu/>

Endeavor 1/31/18-2/5/18

Pioneer 10 leg 1 (24 Mar - 2 Apr)

Pioneer 10 leg 2 (3 Apr - 14 Apr)

IFCB, DAVPR, EIMS [Taylor Crockford, Zoe Sandwith, Ellen Roosen]

Endeavor 7/20/18 – 7/25/18

**Miscellaneous**

Underway pCO2 sensor:

<https://www.km.kongsberg.com/ks/web/nokbg0240.nsf/AllWeb/7870D62A7A6DD9E9C1257EDC00447C6A?OpenDocument>

Pioneer AUV cruise: April 23-28 (Peter Brickley, contact)

Across-shore mission as planned

Latitude of along-shore mission TBD based on frontal location.

**Incubator Platforms**

Smith Lab: incubator footprint is 60x48”, and we’d like it the base of the incubator to be lifted by 24”. We suggest using ¾” plywood and 4 4x4” posts at each corner, with two additional posts of the same size in the center (since this beast when filled with water is a bit heavy). We likely will bring our own plywood and supports as back-up as well.